

New Jersey Mycological Assn.

NJMA News
Vol. XI

February 1981
No. 2

President: Ray Fatto

Editor: Melanie Spock 3.

Circulation: Edythe Krape

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CHEFS COOK 8

Professional chefs Paul Leuthard and Max Meister are scheduled to prepare wild mushrooms at the February 8 meeting. This will be their third year cooking for us and they present an impressive show. Paul is the head chef for the executive dining room of Avon Corporation and Max is Paul's sous chef. The Four Seasons, the Rainbow Room, Plaza One and the Tower Suite are some of the restaurants with which both chefs had been associated. We will get to sample the culinary treats. Anyone having dried or frozen mushrooms that they would like to contribute for the meeting, call Grete Turchick at 201-327-7926. SCEEC auditorium, 2:00 p.m.

FROSTY FUNGI - MARCH 8

The lecture for the March 8 meeting will be on the flora of the tundra (including fungi) and how these life forms cope with the hard environment of the arctic. Anna Gerenday will discuss her kyack trips to Baffin Island and Boothia Peninsula in the arctic and show slides. SCEEC auditorium, 2:00 p.m.

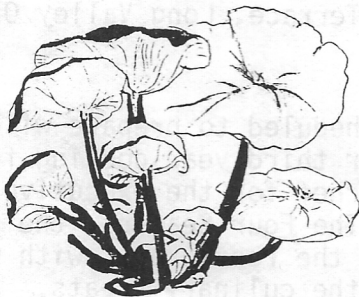
L B M s - APRIL 12

The last lecture this winter will feature LBMs, those numerous, small, dainty, hard to identify mushrooms that everyone finds, but nobody ever wants to bother identifying. Gary Lincoff will tackle the task. At least he will help us discover what they are. Gary is current president of the North American Mycological Association (NAMA) and co-author of *Toxic and Hallucinogenic Mushroom Poisoning*. The meeting will be Sunday, April 12, SCEEC auditorium, 2:00 p.m.

Have you forgotten to pay your 1981 dues? We would like to type the membership roster, and if you don't want to be left out, mail your check today. Individual membership is only \$7.50 and family is \$10.00. Mail your check payable to NJMA to Mrs. Margarete Turchick

Executive Committee Selected

Committee chairpersons have been selected. The executive board will hold a meeting January 18th to plan the coming year. We welcome feedback from members. Any questions, suggestions or complaints pertaining to an area within a committee's responsibility should be referred to the committee chairperson. Keep the following list handy for reference.



	<u>Officers</u>	
President	Ray Fatto	359-5373
Vice President	Anna Gerenday	822-1465
Treasurer	Grete Turchick	827-7926
Secretary	Melanie Spock	356-8935
	<u>Committees</u>	
Books	Glenn Peacock	874-6548
Education	Vic Gambino & [redacted]	398-5622
	[redacted] & [redacted]	609-921-2764
Forays	[redacted]	278-3330
	[redacted]	272-6557
Hospitality	Linda Manailovich	852-1322
Library	Al Northup	538-5153
Membership	Jim Richards	852-1674
Mycoaesthetics	Gertruce Espenscheid	782-7072
Mycophagy	Grete Turchick	827-7926
Newsletter	Melanie Spock	356-8935
Photography	Al Leyenberger	444-3531
Program	Ray Fatto	359-5373
Publicity	Janet Eschenlauer	635-7931
Taxonomy	Dorothy Smullen	647-5740
Toxicology	--	--

NOTES

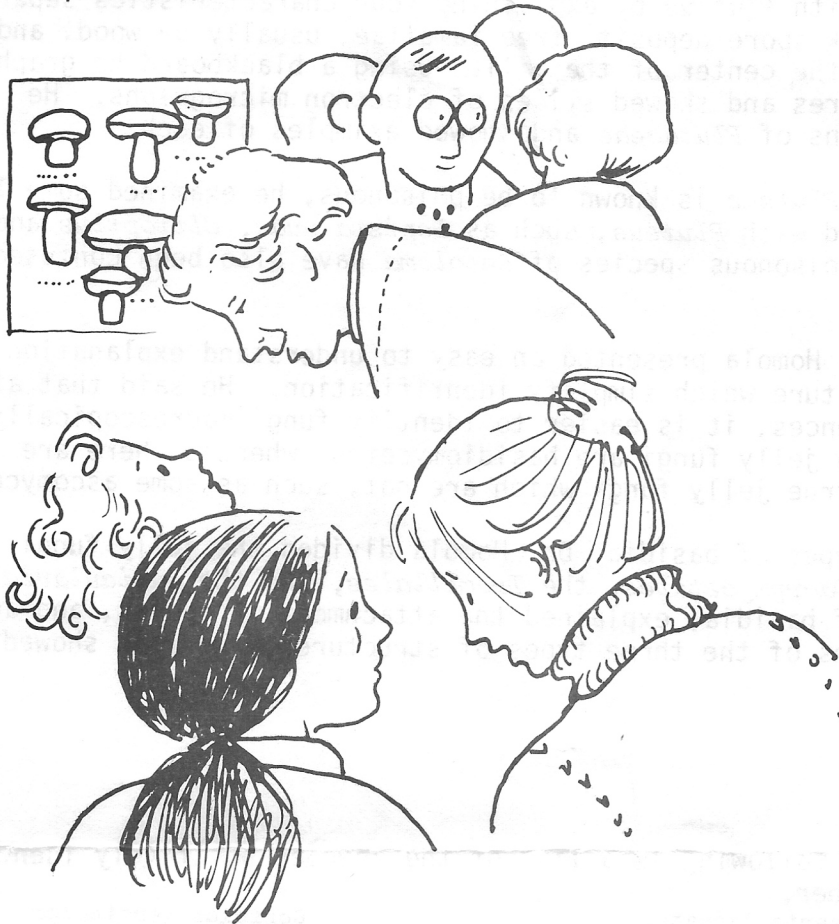
To show our appreciation of Jim Richard's past two years as president, NJMA commissioned Neal MacDonald to paint a mushroom of Jim's choice. At the January meeting we presented Jim with a beautiful watercolor of *Boletus rufocinnemomeus*, a mushroom he frequently finds.

Dr. Clark Rogerson lectured on the dark-spored genera of gilled mushrooms. In one of his usual well-organized lectures, Dr. Rogerson explained the characteristics differentiating the three families of dark-spored mushrooms: *Agaricaceae*, *Coprinaceae*, and *Strophariaceae*. Dividing the families into appropriate genera, he showed slides and explained species in each group which may be

Mentioning Dr. Rene Pomerleau's new book *Flore des Champignons au Quebec*, Dr. Rogerson felt that it was the most comprehensive work for the amateur in the northeast since Kauffman's volumes. Most of the fungi can be found in this area (not only in Quebec) and the book contains reliable first hand information on edibility. The only hitch is that it is written in French.

Dr. Rogerson compiled a key on the dark-spored genera, based on a translation of Pomerleau's book, which he handed out for us to try this season.

Tina Marasmius
Epithets



TAXONOMY

No taxonomy meeting has been scheduled for February. Much cataloguing still needs to be done. Those wishing to work on taxonomy or cataloguing, please call Dorothy Smullen and a day and time can be arranged, 201-647-5740.

CLASSES OFFERED

Brush up before the collecting season starts! Five classes which should appeal to all levels of club membership have been scheduled for the coming year. The classes will all be held on Saturdays at SCEEC. They will be downstairs in classroom #3.

The club is asking for a donation to cover room rental and some classes require a nominal additional expense for course materials.

*I think he's more luridus than badius
if you know what I mean.*

- February 28 - Introduction to Fungi, for beginners, Vic Gambino, 1:30 - 3:00
- March 14 - Identification to Genus, for beginners, Bob Peabody, 11:00 - 3:00 p.m.
- March 28 - Identification to Genus Expanded, Bob Peabody and Dorothy Smullen, 11:00 - 3:30 p.m.
- April 4 - Macroscopic Identification, Gary Lincoff, 1:00 - 4:00 p.m.
- April 11 - Macroscopic Identification, Gary Lincoff, 1:00 - 4:00 p.m.

Vic's class is a basic beginning identification course. It is designed for someone with absolutely no knowledge of fungal types. No complex technical language will be used and all terms will be explained. Slides and handouts will be used. A donation of \$1.00 for the class will cover room rental. To register, call Vic at [REDACTED]

When the details are worked out for the other classes, more information will be in a subsequent newsletter.

Two For One

Dr. Richard Homola's special lecture on December 28th was special in that he treated us to two lectures: one on jelly fungi, as announced, and also one on the genus *Pluteus*. He started off with *Pluteus* by explaining four characteristics separating it from other genera: pink spore deposit, free lamellae, usually on wood, and inverse lamellar trama on the center of the gill. Using a blackboard he graphically explained cellular structures and showed slides of electron micrographs. He discussed the three sections of *Plutaceae* and showed examples of each.

Noting that no species of *Pluteus* is known to be poisonous, he examined some look-alikes that can be confused with *Pluteus*, such as *Lepista nuda*, *Clitopilus* and young *Agaricus* species. Poisonous species of *Entoloma* have also been confused with *pluteus*.

In the "jelly" lecture Dr. Homola presented an easy to understand explanation of basic differences in structure which simplify identification. He said that after seeing microscopic differences, it is easier to identify fungi macroscopically. He explained that the true jelly fungi are basidiomycetes, whereas there are fungi that look like the true jelly fungi which are not, such as some ascomycetes.

Based on differences in types of basidia, Dr. Homola divided the jelly fungi into three major groups: the *Dacrymycetales*, the *Tremellales*, and the *Auriculariales*. He drew the three types of basidia, explained the attachment of spores, and again showed electron micrographs of the three types of structures. He then showed slides of the fungi.

Fair List

Thanks to Anna Gerenday, the following is a list of the species positively identified from the fungus fair in October.

Agaricus campestris	Laccaria laccata	Coriolus versicolor
Amanita gemmata	Lentinus lepideus	Daedalea confragosa
A. muscaria	Mycena alcalina	Fomes annularis
A. virosa	Mycena haematopa	Fomitopsis (Fomes) pinicola
Coprinus micaceous	M. galericulata	Grifola (Polyporus) frondosus
C. ephemerus	Omphalotus olearis	Ischnoderma (Polyporus) resinosum
C. niveus	Panellus stipticus	Oxyporus (Fomes) populinus
C. disseminatus	Panus strigosus	Phellinus ignarius
Entoloma abortivum	Pleurotus ostreatus	Piptoporus betulinus
Lepiota naucina	Gyrodon meruloides	Polyporus albellus
Lepiota cristata	Leccinum scabrum	P. adustus
L. procera	Suillus grevillei	P. hirsutus
Pluteus cervinus	S. granulatus	P. pargamenus
Russula nigricans	S. Americanus	P. sulphureus
Hypoholoma fasciculare	Cantharellula (Cantharellus) umbonata	P. tephroleucus
Pholiota squarrosa-adiposa	Phlebia radiata	Calvatia cyathiformis
P. alnicola	Fistulina hepatica	Lycoperdon perlatum
Stropharia rugoso-annulata	Hericium coralloides	L. pyriforme
S. coronilla	H. ramosum	Crucibulum laeve
Armariella mellea	Ganoderma applanatum	Cyathus striatus
Hygrophoropsis aurantiaca	G. lucidum	Phallus ravenelii
		Scleroderma (Calvatia) cepa

First Class

The executive committee has voted to drop the bulk rate postage permit. All newsletters will now be mailed first class mail, which will eliminate complaints of members receiving newsletters late. With the bulk rate mailing we were able to send up to an ounce for about 8¢ (28¢ first class mail). Since the cost of first class mail may go up, and to save on printing costs, it will be necessary to cut the size of the newsletters.

LEARNING HOW TO DRAW *by GERTRUDE ESPENSCHIED*

You don't have to be an artist to learn how to draw. In 1974 I wrote to Roy Watling asking him to send drawings to an exhibition of drawings of fungi for the 1975 NAMA foray at Dartmouth. He had been listed in a catalog of botanical artists.* He sent back a charming letter begging off, saying he wasn't an artist and his drawings were not suitable.

But in 1976 when his *Identification of the Larger Fungi* was published, I saw his very good drawings that illustrate the book. Roy Watling draws very well indeed.

For that same exhibition, a German chemist Dr. Fritz Wohlfarth sent beautiful Mushroom drawings. When I admired them, he told me he had taught himself to draw out of need for food during and after WW II. He saw mushrooms all around and made very accurate drawings of them to show later to anyone who could identify them - or anyway tell him whether he could eat them. "I'm not an artist" he said. But in making the drawings, he learned to produce admirable watercolor drawings that collectors seek to own. We sold all the drawings he sent.

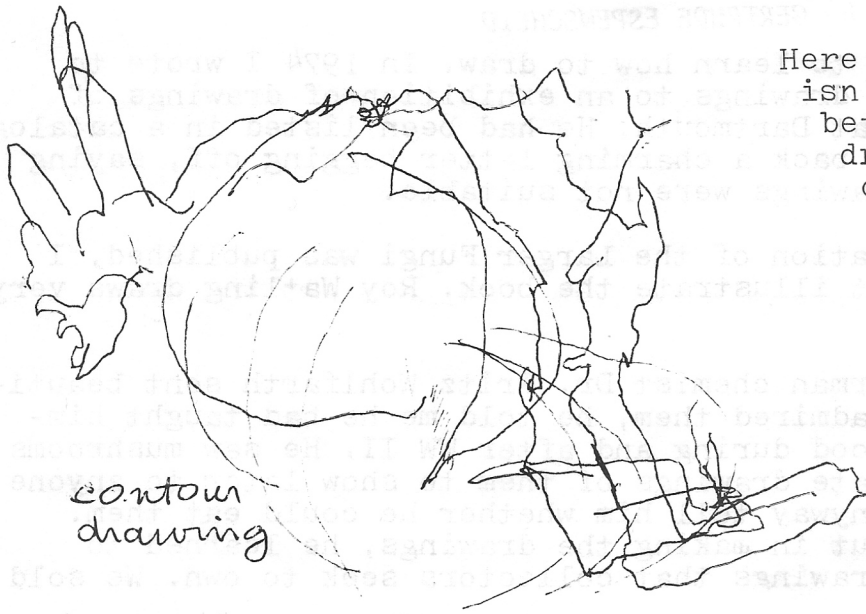
What is it that keeps so many of us from this natural form of expression? It is a big dismal matrix of prejudice and that's all! Drawing is natural. Children do it spontaneously. Anyone with normal vision and intelligence can learn. Drawing is not only a pleasure, but a very useful skill.

There are exercises you can do to get started. I will describe those that were (and are) most helpful to me - three in this article and more in the article to follow. And I will recommend a book that can give you much more help. This first exercise was publicized by Kimon Nikolaides in 1941 in his book *The Natural Way to Draw*. The method is now widely known and used by artists and art teachers. It is called Contour Drawing. Contour Drawing is done without looking at the drawing at all, but looking only at the object (model) being drawn.

Start with something at hand. In mid-winter wild mushrooms are hard to come by, but choose something that has an interesting somewhat complicated shape. I chose a sprouting onion for the example. Tape or tack your paper to your drawing board so it can't slide around. Place the object to be drawn at one side and have the drawing on your other side, out of your vision, so that while you keep your eye on the object, the drawing can not be seen.

Now look at the object and choose a place to begin. Put your pencil point on the paper, and as though your eye and your pencil were one, very slowly move your eye along the contour of the object, with the pencil simultaneously describing that form on the paper. Do not peek at the drawing. The pencil must record every least change in direction. Don't talk to yourself about the form - just sense it and draw it at the same time. If you hear yourself saying in your head, "Now it goes so.." say to yourself, "Be quiet. I am working". Keep your head quiet. Try to feel that the pencil is actually moving over the object - the model. Move all over the object, feeling every rise and fall, drawing every change in direction, until you have drawn everything you can see to draw. You have not looked at the drawing until now.

*Hunt Institute for Botanical Documentation, Pittsburgh, Pa.
"Botanical Art and Illustration 1972-1973"



contour drawing

Here it is - the little gem. Neat, isn't it? Yours will probably be better. We are not after a great drawing. We have had the experience of drawing, and we are learning to coordinate eye and hand.

The next exercise uses Contour Drawing again, but this time, you will look at the drawing when you have lifted the pencil, or just to check as you proceed. Draw only when looking at the object, drawing slowly, and always imagining that the pencil is on the model. This method has been called Modified Contour Drawing. The method will become

very useful to you. Many artists use it throughout their lives. These are good daily exercises. Try to save some quiet time every day for drawing, using the two methods outlined above. After a few days add one more exercise that will help you to keep images in your mind: Memory Drawing.

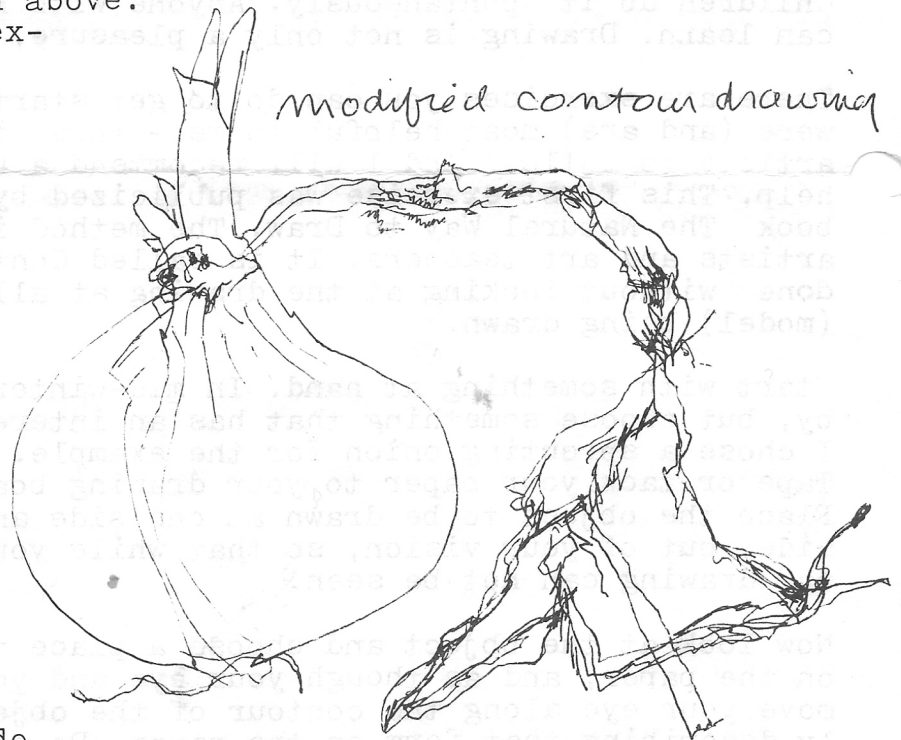
For Memory Drawing take any handy object and study it carefully. Then put it out of sight and draw it.

Put the object back where you had it when you studied it and look at it again. Don't worry about the quality of the drawing. You are learning to keep images in your mind.

Do all three exercises every day.

There is an excellent book on drawing that was published in 1979 - Drawing on the Right Side of the Brain, by Betty Edwards. J.P. Tarcher, publisher. \$8.95 paperback. I strongly recommend it for its splendid self-teaching method. It is full of widely different exercises and examples.

Betty Edwards took the scientific discoveries relating to the two halves of the brain, and adapted her teaching toward developing the right side of the brain- the side that 'images', sees spatially, visualizes - is the artist in us. The left side is the intellectual, the verbal half. Edwards gives exercises to hush the chatty verbalizing left half so the right brain can have quiet to draw. And that's just what you want.



modified contour drawing

Northeast News

Three hundred and ninety different fungi were recorded at the 5th annual northeast foray, despite what were considered very poor collecting conditions. There were 340 Basidiomycetes, 42 Ascomycetes, 3 Deuteromycetes, 1 Phycmycete, and 14 Myxomycetes. Most of the collecting took place in hemlock-hardwood forest, with the latter consisting primarily of sugar maple and white birch. There were some collecting areas that had red oaks and white pines. The lack of moisture was disappointing, however. A single shower in the area, about five days before the foray, broke what had been a month long drought.

More than 285 people, including professional mycologists, attended the foray. The attend-

ing mycologists were: Dr. Tim Baroni, Dr. Howard Bigelow, Dr. Margaret Barr Bigelow, Dr. Roger Goos, Dr. John Haines, Dr. Roy Halling, Dr. Kenneth Harrison, Dr. Richard Homola, Dr. Geoffrey Kibby, Dr. Sam Ristich, Dr. Alexander H. Smith, and Dr. Carl B. Wolfe.

A number of specimens from the collection have been placed in herbaria. They are indicated by the abbreviations following the species names: UMASS = University of Massachusetts, URI = University of Rhode Island, NYS = New York State Museum at Albany, REH = Roy Halling, and ECSC = Eastern Connecticut State College. Inquires may be directed to the foray recorder, Dr. Barry L. Wulff, Department of Biology, Eastern Connecticut State College, Willimantic, Ct. 06226.

BASIDIOMYCETES

Agaricales

AGARICACEAE

Agaricus

- A. arvensis
- A. diminutivus
- A. placomyces
- A. rodmani
- A. silvicola group

AMANITACEAE

Amanita

- A. abrupta
- A. bisporigera
- A. brunnescens
- A. caesarea
- A. cokeri
- A. flavoconia
- A. flavorubescens
- A. frostiana
- A. fulva
- A. gemmata
- A. muscaria
- A. pantherina
- A. porphyria
- A. rubescens
- A. vaginata
- A. virosa (UMASS)

BOLETACEAE

Boletellus

- B. chrysenteroides
- B. russellii (REH)

Boletinellus

- B. merulioides

Boletus

- B. affinis
- B. affinis v. maculosus
- B. badius
- B. bicolor
- B. chrysenteron
- B. edulis
- B. flavus
- B. fraternus
- B. griseus
- B. longicurvipes
- B. luridus
- B. ornatipes (UMASS)
- B. pallidus
- B. rubellus
- B. parasiticus
- B. spadiceus
- B. subglabripes
- B. subtomentosus
- B. subvelutipes
- B. truncatus
- B. vermiculosoides

BOLETACEAE (Con't)

Gyroporus

- G. castaneus
- G. cyanescens

Leccinum

- L. albellum
- L. albogriseum
- L. griseum (UMASS)
- L. holopus
- L. scabrum
- L. snellii

Suillus

- S. americanus
- S. granulatus
- S. grevillei
- S. hirtellus
- S. luteus
- S. pictus

Tylopilus

- T. alboater
- T. chromapes
- T. gracilis
- T. eximius

BOLBITACEAE

Conocybe

- C. lactea

CANTHARELLACEAE

Cantharellus

- C. cibarius
- C. cinnabarinus (UMASS)
- C. ignicola
- C. infundibuliformis
- C. xanthopus

Craterellus

- C. fallax
- C. subfoetidus (UMASS)

Gomphus

- G. floccosus

COPRINACEAE

Coprinus

- C. atramentarius
- C. micaceus
- C. quadrididus

Psathyrella

- P. atomata
- P. candolleana
- P. velutina

CORTINARIACEAE

Cortinarius

- C. armillatus
- C. anomalus
- C. evernius
- C. iodes
- C. splendipes
- C. torvus
- C. violaceus

Crepidotus

- C. applanatus
- C. crocophyllus
- C. mollis

Gymnopilus

- G. spectabilis

Inocybe

- I. caesariata

ENTOLOMATACEAE

Entoloma

- E. grayanum
- E. luteum
- E. madidum

Nolanea

- N. salmonea

HYGROPHORACEAE

Hygrophorus

- H. cantharellus (UMASS)
- H. conicus
- H. deceptivus
- H. fibrillosus
- H. flavescens
- H. marginatus
- H. marginatus v. concolor
- H. miniatus
- H. nitidus
- H. nitratius (UMASS)
- H. pallidus (UMASS)
- H. pratensis (UMASS)
- H. psittacinus
- H. purpureifolius (UMASS)
- H. purus (?)
- H. ruber
- H. squamulosus
- H. unguinosus

LEPIOTACEAE

Lepiota

- L. clypeolaria
- L. cristata
- L. procera
- L. rubrotincta
- L. seminuda v. pussillomyces

PAXILLACEAE

Paxillus

- P. atrotomentosus
- P. involutus

Phylloporus

- P. rhodoxanthus

PLUTEACEAE

Pluteus

- P. admirabilis
- P. auriantoluteum?
- P. aurantiorugosus
- P. cervinus
- P. chrysophaeus
- P. flavofuliginosus
- P. pellitus (UMASS)
- P. salicinus
- P. seticeps
- P. tomentosulus

Volvariella

- V. bombycina

RUSSULACEAE

Lactarius

- L. alnicolor
- L. argillaceifolius
- L. camphoratus (UMASS)
- L. chrysorheus (UMASS)
- L. cinereus
- L. colorascens
- L. deceptivus
- L. fumosus
- L. gerardii
- L. griseus
- L. hygrophoroides
- L. indigo
- L. lignyotus v. canadensis
- L. luteolus
- L. nitidus
- L. payattensis (UMASS)
- L. piperatus (UMASS)
- L. pubescens
- L. pyrogalus
- L. serifluus
- L. sordidus
- L. subdulcis?
- L. subpurpureus
- L. subvellereus
- L. tabidus
- L. vinaceorufescens
- L. volemus

Russula

- R. aeruginea
- R. brevipes
- R. clariflava?
- R. compacta
- R. cyanoxantha
- R. fragilis
- R. heterophylla
- R. laurocerasi
- R. lepida
- R. lutea
- R. mariae
- R. nigricans
- R. obscura
- R. ochroleuca
- R. paludosa
- R. peckii
- R. pectinatoides
- R. puellaris
- R. pusilla
- R. sanguinea
- R. silvicola
- R. variata?

STROBILOMYCETACEAE

Strobilomyces

- S. confusus
- S. floccopus (UMASS)

STROPHARIACEAE

Pholiota

- P. squarrosoides

Psilocybe

- P. caerulipes

Stropharia

- S. hardii (RH)

TRICHOLOMATACEAE

Armillaria

- A. dryina s.s.

Asterophora

- A. lycoperdoides

Baeospora

- B. myriadophylla

Cyptotrama

- C. crysopela

Clitocybe

- C. americana (UMASS)
- C. candicans
- C. dealbata (UMASS)
- C. gibba
- C. hydrogramma
- C. odora
- C. truncicola

Collybia

- C. butyracea
- C. confluens
- C. cylindrospora (RH)
- C. dichrous (R:)
- C. dryophila
- C. luxurians
- C. microspora
- C. maculata
- C. subruda

Flammulina

- F. velutines

Hygrophoropsis

- H. aurantiaca

Laccaria

- L. laccata
- L. ochropurpurea

Lentinellus

- L. cochleatus
- L. ursinus
- L. vulpirus

Marasmius

- M. resinosus
- M. rotula
- M. sicca (RH)
- M. strictipes

Melanoleuca

- M. albiflavida

Mycena

- M. haematopus
- M. leiana
- M. rutilantiformis ?

Oudemansiella

- O. radicata

Panellus

- P. stipticus

Pleurotus

- P. ostreatus
- P. sapidus

Tricholoma

- T. umbrosum (UMASS)

Tricholomopsis

- T. decora
- T. platyphylla

Xeromphalina

- X. campanella

Anhyllorhizaceae

CLAVARIACEAE

Clavaria

- C. fusiformis
- C. rubicundula (UMASS)
- C. vermicularis

Clavariadelphus

- C. truncatus

Clavicornona

- C. pyxidata

Clavulina

- C. amythystina
- C. cinerea
- C. cristata

Physalacia

- P. inflata (RH)

Ramaria

- R. stricta

Ramariopsis

- R. kunzei

CORTICIACEAE

Cryptochaete

- C. rufa

Christiansenia

- C. mycetophila

GANODERMACEAE

Ganoderma

- G. applanatum
- G. tsugae

HYDNACEAE

Climacodon

- C. septentrionale

Dentinum

- D. albidum

Hericium

- H. ramosum

Hydnellum

- H. scrobiculatum

HYMENOCHAETACEAE

Coltrichia

- C. cinnamomea (UMASS)

Hymenochaete

- H. agglutinans (ECSC)

Hydnochaete

- H. olivaceum

POLYPORACEAE

Coriolus

- C. versicolor

Dadaelea

- D. confragosa
- D. unicolor

Favolus

- F. alveolaris

Fomes

- F. fomentarius
- F. ignarius

Laetiporus

- L. sulphureus

Lenzites

- L. betulina

Irpex

- I. lacteus

POLYPORACEAE (Con't)

- Oxyporus
 - O. populinus
- Piptoporus
 - P. betulinus
- Polyporus
 - P. adustus
 - P. conchifer
 - P. cuticularis
 - P. dichrous
 - P. fagicola
 - P. nidulans
 - P. perennis
 - P. pergamenus
 - P. picipes
 - P. radicans
 - P. schweinitzii
 - P. spraguei
 - P. squamosus
 - P. tomentosus
 - P. varius
- Pycnoporus
 - P. cinnabarinus
- Tyromyces
 - T. albellus

STEREACEAE

- Stereum
 - S. complicatum
 - S. fasciatum
 - S. striatum

Lycoperdales

GEASTRACEAE

- Gastrum
 - G. saccatum (ECSC)

LYCOPERDACEAE

- Bovista
 - B. sp. (ECSC)
- Calvatia
 - C. cyathiformis
 - C. rubro-flava (ECSC)
- Lycoperdon
 - L. candidum
 - L. coloratum
 - L. perlatum
 - L. pyriforme

CALOSTOMACEAE

- Calostoma
 - C. cinnabarina

Phallales

PHALLACEAE

- Dictyophora
 - D. duplicata
- Mutinus
 - M. caninus

Nidulariales

NIDULARIACEAE

- Crucibulum
 - C. laeve (URI)

Sclerodermales

SCLERODERMACEAE

- Scleroderma
 - S. areolatum
 - S. arenicola
 - S. citrinum

Tremellales

AURICULARIACEAE

- Phleogena
 - P. faginea

DACRYMYCETACEAE

- Calocera
 - C. cornea (URI, ECSC)
- Dacrymyces
 - D. palmatus

TREMELLACEAE

- Pseudohydnum
 - P. gelatinosum (ECSC)
- Tremella
 - T. concrescens (ECSC)
 - T. reticulata (ECSC)
 - T. tuberculosa (UMASS)
- Tremellodendrom
 - T. schweinitzii (ECSC)

ASCOMYCETES

Heliotales

DERMATIACEAE

- Chlorosplenium chlora

Geoglossaceae

- Microglossum rufum
- Spathularia flavida (ECSC)
- Trichoglossum hirsutum
- Trichoglossum farlowii (ECSC)

LEOTIACEAE

- Bisporella citrina (ECSC)
- Chlorociboria aeruginascens
- Hymenoscyphus fructigena
- Leotia atrovirens
- Leotia lubrica

SCLEROTINIACEAE

- Rutstroemia macrospora (ECSC)

Pezizales

HELVELLACEAE

- Helvella crispa (ECSC)
- Helvella villosa (ECSC)

PEZIZACEAE

- Peziza badia

PYRONEMACEAE

- Humaria hemispherica
- Otidia leporina
- Scutellinia scutellata (ECSC)

SARCOSYPHACEAE

- Sarcoscypha occidentalis

SARCOSTOMACEAE

- Galiella rufa
- Pseudoplectania sp. (ECSC)

Pleosporales

VENTURIACEAE

- Apiosporina morbosa

Sphaeriales

CLAVICHIACEAE

- Cordyceps militaris
- Cordyceps ravenelii (ECSC)
- Cordyceps sp.

DIATRYPACEAE

- Diatrypella betulina (UMASS)
- Diatrypella decorata (UMASS)

HYPOCREACEAE

- Hypocrea avellanea
- Hypocrea gelatinosa
- Hypocrea lactea (ECSC)

NECTRIACEAE

- Hypomyces chrysospermus
- Hypomyces lactifluorum (UMASS)
- Hypomyces luteovirens (UMASS)
- Hypomyces hyalinus (NYS)
- Nectria cinnabarina

SORDARIACEAE

- Lasiosphaeria hispida

XYLARIACEAE

- Daldinia concentrica (URI)
- Hypoxylon fragiforme (ECSC)
- Ustilinia deusta
- Xylaria hypoxylon
- Xylaria polymorpha

DEUTEROMYCETES

- Hormomyces coralloides (ECSC)
- Sepedonium chrysospermum
- Stilbella orbiculare

PHYCOMYCETES

- Spinellus sp. on M. haematopus

MYXOMYCETES

Arcyria

- A. cinerea (ECSC)
- A. denudata (ECSC)

Ceratomyxa

- C. fruticulosa

Fuligo

- F. septica

Hemitrichia

- H. stipitata (ECSC)

Leocarpus

- L. fragilis (ECSC)

Lycogala

- L. epidendrum

Physarum

- P. bivalve
- P. roseum (ECSC)
- P. viride (ECSC)

Reticularia

- R. sp.

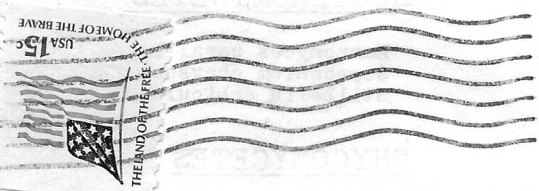
Stemonitis

- S. splendens (ECSC)
- S. sp. (ECSC)

Tubifera

- T. ferruginea (ECSC)

Sue Hopkins: Layout
 Bernice Fatto: Illustrations p. 1, p. 2
 Gertrude Espenscheid: Cartoon
 Dan Graham: Turtle Illustration
 Northeast Foray List: Barry Wulff



New Jersey Mycological Assn.
c/o Ray Fatto, President

Did you ever wonder why "Snapper Turtle Soup" appears on menus, but not "Box Turtle Soup"? Bob Peabody has researched this little problem and found that land turtles - like the box turtle - do not have a liver to eliminate toxins. They can also eat plants and mushrooms which are toxic to humans, but not toxic to the turtle. Therefore, if a land turtle eats a poisonous mushroom, and you eat the turtle, guess what happens?

